

## RhoA/Rho-kinase and vascular diseases: what is the link?

Kenia Pedrosa Nunes · Christine S. Rigsby ·  
R. Clinton Webb

Received: 22 February 2010 / Revised: 7 July 2010 / Accepted: 8 July 2010 / Published online: 29 July 2010  
© Springer Basel AG 2010

**Abstract** RhoA/Rho-kinase pathway plays an important role in many pathological conditions. RhoA participates in the regulation of smooth muscle tone and activates many downstream kinases. The best characterized are the serine/threonine kinase isoforms (Rho-kinase or ROCK), ROCK $\alpha$ /ROCK2 and ROCK $\beta$ /ROCK1. ROCK is necessary for diverse functions such as local blood flow, arterial/pulmonary blood pressure, airway resistance and intestinal peristalsis. ROCK activation permits actin/myosin interactions and smooth muscle cells contraction by maintaining the activity of myosin light-chain kinase, independently of the free cytosolic calcium level. The sensitization of smooth muscle myofilaments to calcium has been implicated in many pathological states, such as hypertension, diabetes, heart attack, stroke, pulmonary hypertension, erectile dysfunction, and cancer. The focus of this review is on the involvement of RhoA/Rho-kinase in diseases. We will briefly describe the ROCK isoforms and the role of RhoA/Rho-kinase in the vasculature, before exploring the most recent findings regarding this pathway and various diseases.

**Keywords** RhoA · Rho-kinase · Cardiovascular diseases · Diabetes · Pulmonary hypertension · Erectile dysfunction · Cancer

### Introduction

Rho-kinase is a serine/threonine protein kinase that contains an N-terminal catalytic kinase domain. It has been identified as the downstream effector of RhoA which mediates calcium ( $\text{Ca}^{+2}$ ) sensitization [1]. RhoA small GTPase (a member of the Rho subfamily within the RAS superfamily of monomeric GTPases) is the molecular switch for various extracellular signals and is implicated in a variety of biological cellular functions, including contraction, migration, adhesion, cell cycle progression, and gene expression. These functions are regulated by RhoA through Rho-kinase or ROCK, one of the best characterized Rho effectors, which exists in two isoforms: ROCK 2 (also called ROCK $\alpha$ ) and ROCK1 (also known as ROCK $\beta$  or p160ROCK) [2, 3]. Rho-kinase is activated not only by RhoA but also by arachidonic acid, which is released from smooth muscle in response to various agonists [4].

Many of the targets for Rho-kinase have been identified, including the myosin-binding subunit (MBS) of myosin phosphatase, ezrin-radixin-moesin (ERM) family, adducin, vimentin (an intermediate filament),  $\text{Na}^+\text{H}^+$  exchanger, and LIM-kinase, which phosphorylates cofilin. Among these, one of the main substrates of Rho-kinase is MLCP (myosin light chain phosphatase), which is physiologically responsible for the dephosphorylation of the light chains of myosin II (MLC<sub>20</sub>) [5]. Thus, MLCP phosphorylation is believed to be a hallmark of Rho-kinase activation [6]. Rho-kinase can also phosphorylate MLC directly [7]. Since the inactivation of MLCP is associated with increased phosphorylation of MLC, the net effect of Rho-kinase activation is consistent with increased phosphorylation of MLC. Phosphorylation or activation of myosin enables the molecular interaction with actin, leading to muscle contraction.

---

K. P. Nunes (✉) · C. S. Rigsby · R. C. Webb  
Department of Physiology, CA3101, School of Medicine,  
Medical College of Georgia, 1120 15th Street, Augusta,  
GA 30912, USA  
e-mail: knunes@mcg.edu; keniapedrosa@gmail.com

Since the contraction of a smooth muscle cell (SMC) occurs through two main mechanisms,  $\text{Ca}^{+2}$  signaling cascades and RhoA/Rho-kinase signaling pathways, Rho-kinase has been shown to be substantially involved in this process. In addition, RhoA/Rho-kinase can alter the  $\text{Ca}^{+2}$  sensitivity of the contractile system [8], and its activation inhibits endothelial nitric oxide synthase (eNOS), thereby altering nitric oxide (NO) production. The impairment of both processes has been shown in human and animal studies to be involved in pathological conditions, mainly vascular diseases and other pathologies, such as hypertension, stroke, vasospasm, atherosclerosis, heart failure, pulmonary hypertension, and more recently, cancer [5, 9–13]. Many of these pathologies demonstrate a common theme: the rapid and dynamic reorganization of the actin cytoskeleton in which Rho-kinase signaling has now emerged as a major switch control.

The RhoA/Rho-kinase pathway has been largely investigated in the last decade, but many aspects regarding this signaling cascade are still unclear. Considering that RhoA mediates important cellular functions and has already been implicated in the regulation of vascular tone, along with inflammation and oxidative stress, the inhibition of this pathway may have significant clinical implications. Some compounds have been studied to inhibit Rho-kinase [14] and have been proposed to have therapeutic benefits regarding multiple diseases. The most widely used experimentally are two non-selective inhibitors, Y27632 and H1077 or fasudil. However, these inhibitors cannot distinguish between ROCK isoforms or the differential mechanisms of ROCK in individual cell components [15], so the precise role of ROCK in the vasculature has thus far been limited by a lack of specific pharmacological inhibitors. Nevertheless, these inhibitors of Rho-kinase have contributed greatly to elucidate altered mechanisms in vascular diseases and have helped to highlight them as therapeutic targets. This review summarizes the current status of this pathway and illustrates its role in multiple diseases, including some speculations on the therapeutic benefits of Rho-kinase inhibitors. The role of RhoA/Rho-kinase in the vasculature, as well as its isoforms and expression, will also be briefly described.

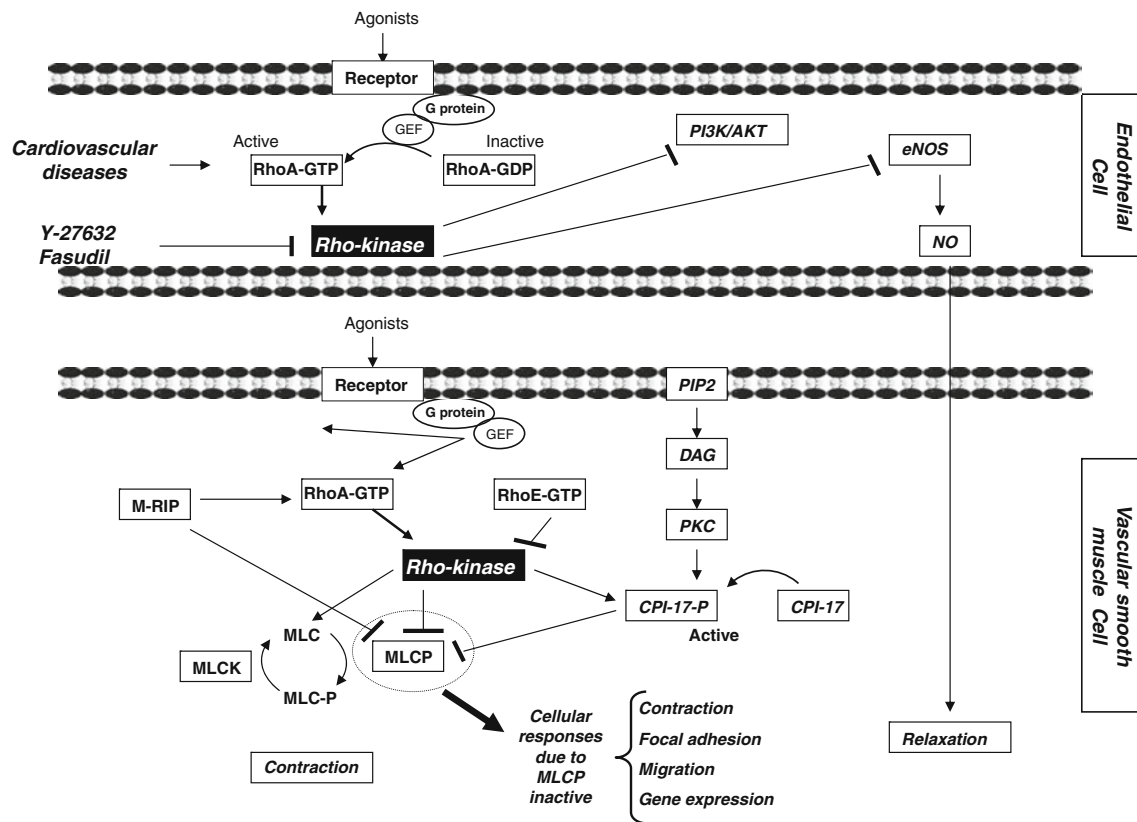
### RhoA/Rho-kinase activity, isoforms, and expression

Rho-family proteins have lipid modifications that target them to cell membranes and they can cycle between GTP- and GDP-bound states [6]. Like other GTP-binding proteins, RhoA exhibits both GDP/GTP-binding activity and GTPase activity, and functions as a molecular switch, cycling between a GDP-bound inactive state (GDP-Rho) and a GTP-bound active state (GTP-Rho). The activity of

RhoA is cyclically regulated [6]. When cells are stimulated with various agonists, GDP-Rho is converted to GTP-Rho through the action of guanine nucleotide exchange factors (GEFs) that stimulate the GTP–GDP exchange reaction. GTP-Rho is then targeted to the cell membrane through its C-terminal geranyl-geranylated tail and interacts with its specific targets (Fig. 1). In resting cells, Rho GDP dissociation inhibitor (Rho GDI) binds to GDP-Rho and extracts GDP-Rho from the membrane to the cytosol [16–18].

Rho-kinase is a widespread and evolutionary conserved downstream effector of RhoA, as well other GTPases, RhoB and RhoC, which are all potential activators. However, there are inhibitory GTPases such as RhoE that directly bind to ROCK and block its kinase activity. ROCK1-induced stress fiber formation is inhibited by RhoE bound to the amino-terminal region [19, 20]. Differently, Rho-A activates ROCK1 through interaction with carboxy-terminal site. RhoE and RhoA are not able to bind ROCK1 simultaneously [21], and RhoE phosphorylation events seem to antagonize RhoA-induced stress fiber assembly [20]. ROCK activation has also been suggested to occur directly by lipids, such as sphingosylphosphorylcholine (SPC) [22]. Rho-kinase was initially identified as a GTP-Rho-binding protein from bovine brain by affinity column chromatography on matrix-bound GTP-Rho [23]. There are two isoforms of Rho-kinase, ROCK1 and ROCK2, that share overall 65% homology at the amino-acid level and 92% homology in their kinase domains [3]. The tissue distribution of ROCK1 and 2 is similar, and relatively few studies have outlined the isoform specific roles of ROCK. Although ROCK1 rather than ROCK2 was suggested as important for stress fiber formation, ROCK2 acts as a counterbalance in the regulation of the microfilament bundle and focal adhesion site [24]. ROCK1, but not ROCK2, is sensitive to caspase 3-mediated cleavage in apoptotic cells [25]. Overexpression of both isoforms increases MLC phosphorylation [26] and, consequently, ROCK isoforms lose their specificity when overexpressed [24]. Analysis of the ROCK2-deficient mouse suggested that there is no compensation for the loss of ROCK2 by ROCK1 [27]. However, in vascular smooth muscle cells (VSMC), silencing of each ROCK isoform leads to upregulation of the other isoform, suggesting that the expression level of the ROCK isoforms is tightly controlled and interrelated. When both ROCK isoforms are silenced, this leads to reduced myosin binding subunit (MBS) and MLC phosphorylation [26].

ROCK isoforms are expressed in invertebrates such as *Caenorhabditis elegans*, *Drosophila*, and mosquito, and vertebrates such as zebrafish, *Xenopus*, chicken, mouse, rat, and human [28]. ROCK1 and ROCK2 are ubiquitously expressed in mouse tissues from youngest embryonic development to adulthood. Evaluation of expression in the



**Fig. 1** The RhoA/Rho-kinase pathway. Agonist triggers RhoA (RhoA-GTP) by stimulation of G-protein-coupled receptors leading to Rho-kinase activation. In stimulated cells, RhoA-GDP is converted in RhoA-GTP through the action of GEF. The activated Rho-kinase subsequently inhibits the eNOS and PI3 K/AKT pathways, impairing NO released from endothelial cells. There is also inhibition of MLCP activity in vascular smooth muscle cells leading to contraction. RhoA/Rho-kinase activation is common in cardiovascular diseases. CPI-17 is activated by Rho-kinase and PKC in vascular smooth muscle cells, resulting in MLCP inhibition. The complex M-RIP protein/RhoA/Rho-kinase also inactivates MLCP, which alters the contractile

process and many cellular responses as cited in the figure. Y-27632, fasudil and activated RhoE are Rho-kinase inhibitors. ROCK-induced contraction is inhibited by RhoE bound to the amino-terminal region, whereas Rho-A activates ROCK1 through interaction with carboxy-terminal site. Rho-kinase, representing two isoforms of ROCK1 and ROCK2. GEF Guanine nucleotide exchange factor, MLCP myosin light chain phosphatase, MLCK myosin light chain kinase, MLC myosin light chain, MP-RIP myosin phosphatase-rho interaction protein, PIP2 phosphatidylinositol 4,5-bisphosphate, DAG diacylglycerol, PI3K phosphatidylinositol-3-kinase

mouse indicates that ROCK1 mRNA is preferentially expressed in lung, liver, spleen, kidney, and testis, whereas ROCK2 mRNA is highly expressed in the heart and brain [29, 30]. Recently, it has been demonstrated that there are direct increases in neural ROCK2 expression in ischemic brain tissue [31]. Immunolocalization and cell fractionation studies have shown that ROCK2 is distributed mainly in cytoplasm [23, 32]. In contrast, little is known regarding the intracellular localization of ROCK1, which may be colocalized to centrosomes [33] and in the Golgi complex, together with RhoE [19].

### RhoA/Rho-kinase in the vasculature

A perfect balance between contraction and relaxation of smooth muscle is critical in maintaining many biological

functions and disruptions in this dynamic process can result in various pathologies. Vascular smooth muscle tone plays a fundamental role in regulating blood pressure, blood flow, capillary permeability, and other cardiovascular functions. The Rho-kinase pathway is intrinsically involved with the process of smooth muscle contraction [24] and has been shown to be a regulated determinant of numerous cellular processes [4]. The  $\text{Ca}^{2+}$  sensitivity of smooth muscle reflects the ratio of activities of MLCP to myosin light-chain kinase (MLCK), resulting in contraction or relaxation. Since the main substrate of Rho-kinase is MLCP, which is physiologically responsible for the dephosphorylation of MLC, this implicates Rho-kinase in mediating  $\text{Ca}^{2+}$  sensitivity inside the cell, ultimately leading to contraction by inhibition of MLCP activity.

It is now widely accepted that MLCK and the RhoA/Rho-kinase pathway are two major cellular targets for

regulating  $\text{Ca}^{2+}$  sensitivity of myosin II, and they generally operate in parallel. RhoA is an important messenger of  $\text{Ca}^{2+}$  sensitization, and agonists can activate RhoA through numerous G protein-coupled receptors:  $\alpha$ -adrenergic, muscarinic, purinergic, endothelin, prostanoid, oxytocin, epidermal growth factor, ephrin, semaphorin, angiotensin II, and Edg lysophospholipid receptors [6, 34]. Activation of RhoA and also phospholipase C (PLC) induces inositol 1,4,5-triphosphate ( $\text{IP}_3$ ) production and  $\text{Ca}^{2+}$  release from the sarcoplasmic/endoplasmic reticulum, and increased  $\text{Ca}^{2+}$  influx through receptor-operated or voltage-gated channels, and inhibits maxi-K potassium channels resulting in depolarization. Also, the linkage between  $\text{Ca}^{2+}$ -calmodulin (Ca-CAM) activates the MLCK, which in turn leads to increased phosphorylation of MLC, promoting the actin filament cross-linking activity of myosin II, and resulting in contraction [35].

PLC activation catalyzes the formation of diacylglycerol (DAG), a second messenger, leading to protein kinase C (PKC) activation, which phosphorylates specific target proteins. There are several isozymes of PKC and each has a tissue-specific role [36]. However, in many cases, PKC has contraction-promoting effects, such as phosphorylation of different kinases including Rho-kinase and others such as MLCK, ERK, calmodulin-dependent protein kinase II, transporters, and various ion channels [37]. So, PKC participates in the contractile response mainly by directly activating MLCK and, indirectly, by activating Rho-kinase.

Rho-kinase was first identified to phosphorylate the myosin-binding subunit (MBS) of MLCP, named myosin phosphatase targeting (MYPT1) subunit, inhibiting its activity. Later, many kinases were found to phosphorylate MYPT1, thus promoting the phosphorylated state of myosin. Within MYPT1 or MBS, which are often used synonymously in the literature, the major sites of phosphorylation by Rho-kinase have been identified as Ser849/854, Thr850/855, and Thr695/697. The Ser849/854 site is specifically phosphorylated by Rho-kinase, but the main site involved in the inhibition of MLCP activity is Thr695/697 [17]. However, recently, Thr850/855 has been implicated as the major ROCK phosphorylation site, whereas Thr695/697 is thought to be phosphorylated by other kinases [38]. The specific phosphorylation sites on ROCK and their activities still need to be elucidated.

Similar to MYPT1, the phosphorylation of the small protein CPI-17, a phosphorylation-dependent inhibitory protein of MLCP, can be catalyzed by multiple kinases including Rho-kinase. Specifically, inhibition of the activity of MLCP involves Rho-kinase by means of phosphorylation of either MYPT1 at Thr850/855 of MLCP or the protein CPI-17, which inhibits the catalytic domain of MLCP [39, 40]. CPI-17 can be phosphorylated by PKC as well as by Rho-kinase (Fig. 1). Therefore, CPI-17 can be

seen as a potential mediator of  $\text{Ca}^{2+}$  sensitization which is independent of MYPT1 phosphorylation.

New members of the MLCP complex have been characterized myosin phosphatase-rho interacting protein, M-RIP, and its murine homolog MP-RIP or p116<sup>RIP</sup>. Both are cytoskeletal scaffold proteins that bind directly to both RhoA and MBS, targeting MLCP via different mechanisms [41, 42]. One targeting function of M-RIP is to localize the MLCP complex to the actinomyosin contractile filament to dephosphorylate myosin leading to inactivation of MLCP activity [42, Riddick]. However, it has been described that the binding of MP-RIP (or p116<sup>RIP</sup>) to MBS activates MLCP activity. The elimination of MP-RIP by MP-RIP-specific siRNA consistently increased MLC<sub>20</sub> phosphorylation [41, 42]. It is postulated that MP-RIP contributes to the decrease in myosin phosphorylation via activation of the myosin dephosphorylation activity of MLCP and the inactivation of the RhoA pathway [42]. Additionally, since ROCK and MP-RIP bind separate domains of MBS, a model where RhoA bound to MP-RIP and ROCK bound to MBS are brought into proximity by MP-RIP/MBS binding has been suggested [26]. The same authors describe that both isoforms of ROCK can bind to MBS, but ROCK1 interacts with MBS twice as much as ROCK2 [26]. Also, in VSMC, ROCK2 is the predominant isoform that regulates contractility and recent studies demonstrated that ROCK2 regulates force production, as well as contraction [26].

Since the RhoA/Rho-kinase pathway is so heavily involved in the cytoskeletal function of the vasculature, it would be expected that there is a link between this pathway and many vascular diseases. However, the involvement of RhoA/Rho-kinase with the cytoskeleton does not just limit it to diseases of the vasculature, as this review will highlight. Nevertheless, many processes regarding the exact role of RhoA/Rho-kinase activation need to be better understood, as well as the consequences of its dysregulation in different parts of the body. This will lead to a total comprehension of this pathway and its involvement in several pathologies.

## Connections between Rho-kinase and diseases

The small GTPase RhoA and its target, Rho-kinase, are involved in the sequence of events which stimulates vascular smooth muscle contraction, stress fiber formation, cell migration, and, indirectly, blood pressure regulation. In this way, RhoA/Rho-kinase activation has significant effects on various cardiovascular diseases, mainly arterial hypertension [43], atherosclerosis [44], heart attack [45], stroke [46], and others such as coronary vasospasm [47], myocardial hypertrophy [48], myocardial ischemia-reperfusion injury [49], and vascular remodeling [50].

Compounds that specifically inhibit this signaling pathway can offer clinical benefits regarding the treatment of these diseases, as well as contributing pharmacological tools for vascular studies.

Accumulating evidence indicates that endothelial nitric oxide synthase (eNOS), which is an important mediator of vascular function, is regulated by the RhoA/Rho-kinase pathway (Fig. 1) [51, 52]. For example, dominant-negative mutants of RhoA or inhibitors of ROCK have been shown to increase eNOS expression [46, 52]. Since eNOS is protective in the vasculature, the RhoA/Rho-kinase pathway has been suggested to play a critical pathophysiological role in several aspects of cardiovascular disease. Also, there is increasing evidence that eNOS activity could be regulated in part through association with various protein kinases [53, 54]. For example, inhibition of RhoA or ROCK isoforms leads to the rapid activation of the lipid kinase phosphatidylinositol-3-kinase (PI3 K)/Akt pathway and phosphorylation of eNOS [51, 52]. Studies also indicate that ROCK isoforms are activated in patients with a cardiovascular disorder or with associated risk factors [55, 56]. Furthermore, RhoA mRNA expression and activity is increased in aortas from aged rats, suggesting a role of RhoA in the development of age-related cardiovascular disease [57].

Vascular remodeling is an important component of vascular diseases and may be associated with RhoA/Rho-kinase signaling. Vascular remodeling occurs during normal development and participates in various physiological processes. However, structural changes to the vasculature can be pathologic as well as adaptive, leading to arterial disease development, which can contribute to cardiovascular dysfunctions such as hypertension and atherosclerosis. Angiotensin II (Ang II) is a potent growth factor involved in arterial wall homeostasis. Cardiac inflammation via activation of the cardiac Ang II system is suggested to play a role in cardiac remodeling [58]. Since some of the cytoskeletal changes that occur in vascular remodeling, specifically in VSMCs, are induced by Ang II [59], Rho-kinase has been directly linked to pathologic vascular remodeling through evidence suggesting that Ang II activates the Rho-kinase pathway and regulates the cytoskeleton [59–61].

Rho-kinase has also been linked with non-cardiovascular diseases such as diabetes, renal injury, erectile dysfunction, pulmonary hypertension, and cancer. Furthermore, RhoA/Rho-kinase has emerged as a pathway involved in some immuno-pathologies. For example, Rho-kinase has an important function in human immunodeficiency virus (HIV)-1-mediated disruption of the integrity of the blood–brain barrier [62]. Tat is a protein produced and released by HIV-1-infected cells, and circulating Tat can be detected in HIV-1-infected patients [63]. Tat, acting via intact lipid rafts, activates RhoA/Rho-kinase, leading to inhibition of MLCP and increased phosphorylation of

MLC. This suggests that Tat-induced RhoA activation may constitute an early signaling mechanism leading to upregulation of efflux transporters on the brain endothelium, thus limiting antiretroviral drug penetration into CNS [62]. Many of these diseases related to Rho-kinase activation presented with damaged vasculature. The involvement of RhoA/Rho-kinase in these pathologies is clear due to an increasing number of experimental studies, along with the evidence that ROCK inhibitors can be used to treat some of those diseases [13, 64, 65]. Nevertheless, there are many points regarding the precise participation of RhoA/Rho-kinase in these disorders that remain unclear. We will explain some of the pathologies that have been shown to be associated with Rho-kinase activation and present the most recent published data.

## Hypertension

Arterial hypertension is one of the most common cardiovascular disorders characterized by altered vascular tone and increased vascular contractility resulting in high blood pressure [66, 67]. It is accompanied by proliferation, migration of VSMCs, and varying levels of inflammation of the arterial wall, processes that together constitute vascular remodeling [58]. The Rho-kinase pathway plays a crucial role in the regulation of arterial blood pressure [68]. The role of Rho-kinase signaling in arterial hypertension was first recognized in 1997 [69]. In that study, a Rho-kinase inhibitor was observed to reduce arterial blood pressure in three experimental models of hypertension. In addition, Rho-kinase could also regulate blood flow via direct effects on the central nervous system [70, 71] or indirectly through negative effects on eNOS expression and activity [72, 73]. The Rho-kinase pathway is increased in spontaneously hypertensive rats [74, 75] and hypertensive patients [68]. This pathway has been frequently investigated since many studies demonstrated that Rho-kinase inhibitors, such as Y-27632 and fasudil, could be potential tools to treat hypertension and atherosclerosis, as well as other cardiovascular diseases [76]. Recently, a novel and potent selective Rho-kinase inhibitor, SAR407899, with promising antihypertensive activity, has been demonstrated to have a superior effect to that of fasudil and Y-27632 [77]. However, it is still a long way away from being used to treat hypertension clinically.

## Atherosclerosis

Rho-kinase also contributes to the development of atherosclerosis and vascular inflammation [78]. Atherosclerosis is characterized by the cross-talk between excessive inflammation and lipid accumulation. Selective Rho-kinase inhibitors lead to upregulation of eNOS, decreased vascular



inflammation, and reduced atherosclerotic plaque formation [44]. In addition, experiments using ROCK1 (−/−) mice showed that ROCK1 in bone marrow-derived macrophages is critical to the development of atherosclerosis, in part by mediating foam cell formation and macrophage chemotaxis [79]. Recently, it has been suggested that statins, or HMG-CoA reductase inhibitors, which are accepted as first line agents for the treatment of hyperlipidemia to reduce the risk of adverse cardiovascular events [80], induced vascular benefits similar of selective RhoA/Rho-kinase inhibitors in vitro [81, 82], exerting an anti-atherogenic effect that is independent of cholesterol reduction. This effect was observed in cultured VSMCs from both animal models and human samples [83, 84]. Furthermore, in the apolipoprotein E knockout mice, a model of accelerated atherosclerosis, lesion development was inhibited by Y-27632 and was associated with inhibition of ERM, a target protein of Rho-kinase [85].

### Stroke

Rho-kinase pathway activation has been observed in various disorders of the central nervous system [86] and it seems important in the pathogenesis of several cerebral vascular diseases, such as stroke [46, 87] and cerebral vasospasm [47, 88]. Recent studies showed that Rho-kinase is directly involved in neuronal damage that occurs during a stroke. Rho-kinase plays an important role in neuronal apoptosis and in the execution phase of apoptosis [89]. The Rho-kinase inhibitor, fasudil, protects the brain tissue against ischemic damage in vitro [46, 90, 91] and the ischemia-induced delayed neuronal death when the treatment was started 24 h after induction of ischemia [90]. Moreover, fasudil was reported to prevent ischemic neuronal damage in vivo by increasing cerebral blood flow through upregulation of eNOS and decreasing the inflammatory response [31, 46], which is achieved by an inhibition of neutrophil migration [92]. This is supported by studies showing fasudil was not effective against induction of transient ischemia and reperfusion in eNOS knockout mice [46]. Recently, it has been described that this drug triggers proliferation and differentiation of adult neural stem cells at the subventricular zone in mice following hypoxia/reoxygenation injury, suggesting a direct effect of fasudil in neurons [93].

### Heart failure

Rho-kinase is involved in the regulation of myofibrillar  $\text{Ca}^{+2}$  sensitivity in cardiac muscle [94] and contributes to irreversible myocardial damage [45]. Rho-kinase is also involved in the pathogenesis of cardiovascular remodeling [95], and its inhibition plays a significant role in treatment

of the failing heart [96] by limiting infarct size [97], which is the major contributor to the development of heart failure [45]. The cardioprotective effect of Rho-kinase inhibition involves PI3 K/AKT and NOS activation [49, 52]. However, Rho-kinase inhibitor compounds need to be evaluated more closely for their efficacy during varying index ischemia periods, a wide dose range, and in vivo animal models mimicking the clinical setting [98]. Recently, it has been suggested that statins, specifically pitavastatin, could improve cardiac function and remodeling via eNOS production associated with the Rho-kinase pathway [99]. This could be explained using data showing that statins lower the intracellular levels of various proteins, such as RhoA [100]. In addition, statins block the activity of RhoA and prevent the activation of ROCK [101], which can regulate eNOS mRNA stability [51].

### Diabetes

Type II diabetes is often associated with a collection of abnormalities including obesity, hypertension, various vasculopathies associated with the Rho-kinase pathway, and insulin resistance [102]. Impaired endothelium-dependent relaxation is a consistent finding in blood vessels from diabetic animals and patients [103, 104]. An increasing body of evidence intensifies the idea that the injured endothelium in diabetes is due, at least in part, to alterations in RhoA/Rho-kinase signaling. Increased vascular permeability is also a major characteristic of diabetic vasculopathy, and, in this condition, evidence indicates that advanced glycation end products (AGEs) activate RAGE, a major receptor for AGEs, and modulate various cell functions by multiple pathways, including Rho-kinase signaling [105]. RhoA and RAGE can unexpectedly form a complex called RhoA/RAGE, which has been suggested to induce Rho-kinase activation, resulting in reorganization of the actin cytoskeleton, leading to endothelial cell hyperpermeability in diabetes [106]. In the renal cortex of the streptozotocin (STZ)-induced diabetic rat, RhoA is highly expressed suggesting its involvement in diabetic renal injury [107]. The kidney is a major site of diabetic microvascular complications leading to renal failure. In kidney mesangial cells, which upregulate matrix protein synthesis in response to high glucose, it has been shown that RhoA/Rho-kinase is required for the fibrotic effects of high glucose in diabetes induced by STZ [108]. Also, activation of RhoA and Rho-kinase is greater in aortas from diabetic mice compared with non-diabetic mice [109]. These studies demonstrate an important role for the RhoA/Rho-kinase pathway in the pathogenesis of diabetic renal disease.

Rho-kinase is also responsible for the impairment of insulin signaling in Zucker obese rats, and its inhibition

corrected glucose levels [110, 111]. RhoA/Rho-kinase inhibition improved the symptoms of diabetic nephropathy reinforcing the participation of this pathway in diabetes complications [112]. However, chronic treatment of obese db/db mice with fasudil was reported to have no effect on blood glucose levels [112]. In contrast, in normal mice, acute treatment with Y-27632 causes insulin resistance *in vivo* [113]. Since ROCK1-deficient mice exhibited systemic insulin resistance via impaired insulin signaling in skeletal muscle, it has been suggested that ROCK1 can affect glucose homeostasis and insulin sensitivity [114]. In alloxan-induced diabetic rabbits and STZ-induced diabetic rats, the ROCK1 gene and protein were upregulated in penile tissue [115, 116]. The involvement of Rho-kinase in diabetes and its complications is becoming clear due to the increased amount of results showing upregulation of the Rho-kinase pathway in diabetic models, both *in vivo* and *in vitro*. The possibility of using Rho-kinase inhibitors to prevent progression of some diabetic problems, such as diabetic nephropathy, has been suggested as a promising novel therapy [108]. However, it is necessary to first clarify all the implications of Rho-kinase signaling alterations during the development of diabetes before employing this therapy in these patients.

## Cancer

Recently, it is becoming clear that the Rho proteins play an important role in several aspects of cancer development, and each member of the Rho family (RhoA, RhoB, and RhoC) may be engaged at a different level at various tumor progression stages [117–119]. One of the first studies connecting Rho and malignancy showed that RhoA and a related molecule, Rac2, are overexpressed in head and neck squamous cell cancers [120]. Since tumor cell invasion and metastasis require a complex cascade of events, including changes in the cytoskeleton, it is predictable that the RhoA/Rho-kinase pathway would be involved in cancer development [119]. Also, Rho-GTPases could be early markers for tumor progression [118]. There are studies suggesting that Rho-kinase inhibitors would be useful to prevent cancer progression [121–123]. Nevertheless, some of these studies indicate that reduced Rho-protein function contributes to the morphological changes observed in tumor cells. This elevates the risk that inhibition of Rho proteins might promote a more aggressive tumor phenotype [117]. It has been reported that RhoA/ROCK is involved in progression of human gastric cancer [124]. This kind of cancer displays high expression of RhoA, which has been correlated with aggressive metastasis. In addition, gastric cancer cell invasion can be induced via activation of the RhoA/ROCK pathway by IL-6. Then, RhoA expression has been suggested useful as a prognostic factor in patients with

gastric adenocarcinoma [124]. In addition, active RhoA has oncogenic potential which promotes the invasiveness of rat hepatoma cells via the activation of the endogenous RhoA pathway [125]. Also, it was demonstrated that cells transformed by Rho oncogenes do have metastatic potential *in vivo* [126]. RhoA and RhoC siRNA gene therapy, mediated by adenovirus, has been suggested as useful for inhibition of growth and invasion of human gastric carcinoma by the PI3/Akt pathway [127]. However, it has been suggested that RhoC is most involved in the cancer process, mainly in metastasis formation, more so than RhoA and RhoB [128].

There is increasing evidence regarding the role of Rho-kinase in cell proliferation and survival. The requirement for survival signals to prevent apoptosis in normal cells is reduced in tumor cells. Rho family proteins including RhoA have been described in both pro- and anti-apoptotic signaling [129]. In studies involving cancer tissues and cell lines, it was reported that there was a direct relationship between cell proliferation and the level of ROCK2 expression in both malignant and nonmalignant cells [130, 131]. Besides cell proliferation, Rho-kinase has been implicated in the drug resistance observed in liver cancer patients [132]. In ovarian cancer, which is very invasive, the lysophosphatidic acid (LPA)/RhoA/Rho-kinase pathway is intimately involved in the course of ovarian cancer progression, and fasudil administration attenuated the invasiveness of the cells by inhibition of this pathway [122]. Pharmacological modulators of several steps of the Rho-kinase signaling pathway have been investigated, and a new chemical compound, CCG-1423, has demonstrated inhibition downstream of RhoA, which can be used as a pharmacological tool to disrupt the Rho-kinase pathway in cancer [133].

## Pulmonary hypertension

Convincing evidence indicates that Rho-kinase is also involved in pulmonary hypertension (PH) [10, 64, 134, 135], a disease characterized by progressive elevation of pulmonary arterial pressure and vascular resistance due to pulmonary vasoconstriction and vessel remodeling, as well as increased inflammation [136, 137]. In isolated human lung tissue from PH patients, Rho-kinase expression and reactivity were significantly increased compared with controls, and relevant correlations were observed between Rho-kinase activity and the severity and duration of PH [138]. Also, in patients with moderate PH, treatment with intravenous fasudil had beneficial effects, such as decreased pulmonary artery blood pressure and pulmonary vascular resistance [139]. Additionally, PH patients that did not respond to oxygen inhalation, NO inhalation, or nifedipine, 30-min intravenous fasudil treatment

significantly decreased elevated pulmonary vascular resistance [134]. Also, fasudil inhalation for 10 min significantly reduced mean pulmonary arterial pressure in patients with PH [140]. Rho-kinase signaling inhibitors have been suggested as a viable therapeutic agent for PH treatment. However, there is no study showing the long-term effects of Rho-kinase inhibitor administration, such as fasudil, in patients with severe PH, even though, in both human and animals, it is reasonable to argue that Rho-kinase has an important role in PH due to its involvement in the sustained vasoconstriction, vascular remodeling, and inflammation that occurs in this pathology. Accumulating evidence from several laboratories indicates that RhoA/Rho-kinase signaling plays an important role in the pathogenesis of many experimental models of PH, including chronic hypoxia [141–143], monocrotaline [143, 144], VEGF receptor inhibition [137], and mild hypoxia-induced PH in neonatal fawn-hooded rats [145]. Furthermore, Rho-kinase signaling mediated vasoconstriction in severe occlusive PH in rats [137], and small pulmonary arteries exhibit Rho-kinase-dependent increases in myogenic tone in chronic hypoxic PH [146].

In mouse models of PH, treatment with Y-27632 decreased the muscularization of distal pulmonary arteries and upregulated eNOS expression [147]. The beneficial effect of sildenafil on PH is mediated, at least in part, by the inhibition of the RhoA/Rho-kinase pathway [141]. It has been demonstrated that serotonin (5-HT), also synthesized by pulmonary endothelial cells, is internalized in SMCs through the 5-HTT (5-HT transporter) and is linked to RhoA by intracellular type 2 transglutaminase, leading to constitutive RhoA activation [148]. Recent studies showed that enhancement of RhoA and Rho-kinase activities in PH is associated with increased RhoA serotonylation, suggesting direct involvement of 5TT/RhoA/Rho-kinase in proliferation of pulmonary artery smooth muscle cells (PA-SMCs) and platelets, during PH progression [149]. Finally, in addition to atherosclerosis, research indicates that statins improve PH in several rat models. New studies have supplied evidence that reversal of hypoxic PH by simvastatin decreased lung expression and activity of both ROCK1 and ROCK2 [150].

### Erectile dysfunction

Another ailment that has been associated with the involvement of RhoA/Rho-kinase in its pathologic mechanism is erectile dysfunction (ED). The penis becomes erect with the relaxation of arteriolar and sinusoidal smooth muscle. NO produced by eNOS [151, 152] is fundamental for smooth muscle relaxation and, consequently, normal erectile function [153]. Since the main function of Rho-kinase is the regulation of smooth muscle tone [154], the

upregulation of the Rho-kinase pathway increases cavernosal smooth muscle contraction, leading to ED [155, 156]. In addition, RhoA/Rho-kinase-mediated  $\text{Ca}^{+2}$  sensitization has an important role in the regulation of corpora smooth muscle tone and keeps the penis in the flaccid state [157]. Further, in the absence of arousal, the penis remains in the non-erect state by cavernosal vasoconstriction induced mainly by norepinephrine and endothelin 1 (ET-1), which are Rho-kinase-mediated responses [115, 158].

ED is associated with cardiovascular diseases, mainly hypertension [159] and other diseases, such as diabetes [116] and hypogonadism [160]. Cardiovascular diseases, mainly atherosclerosis and hypertension, have been considered a risk factor for ED [161]. In spontaneously hypertensive rats (SHR) and DOCA-salt rats (a mineralocorticoid-dependent hypertensive rat), increased activity of the RhoA/Rho-kinase pathway was observed, and could be the cause of ED in these hypertensive models [158]. However, over the past few years, many studies have suggested that ED may be an early marker in the development of cardiovascular pathologies [162], reinforcing the involvement of Rho-kinase activity in these diseases.

In corpus cavernosum tissue from diabetic rats, it was observed that there was an enhancement of Rho-kinase activity [115]. In streptozotocin-induced diabetic rats, in vivo injection of Y-27632 into the penis increases intracavernosal blood pressure, membrane-fraction RhoA protein expression, and MYPT1 phosphorylation levels, supporting observations that alterations in the Rho-kinase pathway occur in the diabetic rat penis [116]. Conversely, in diabetic mice, even though acetylcholine-induced corpus cavernosal tissue relaxation and erectile function are considerably less than in non-diabetic mice, Rho-kinase inhibitors produce a similar relaxation response, suggesting that RhoA/Rho-kinase signaling may not be altered in the diabetic mouse penis [163]. Regarding hypogonadism-associated ED, it has been suggested that upregulation of the RhoA/Rho-kinase pathway in the penis may be an associated mechanism [164]. The same work demonstrated that testosterone replacement restored erectile function and reduced RhoA and Rho-kinase protein expression in castrated rats, although the expression levels were still significantly increased compared with intact rats. These results could be explained due to multi-factorial effects of testosterone, including NO stimulation [164]. In a castrated rat model of ED, it has been demonstrated that Rho-kinase inhibition was able to improve androgen ablation-induced ED [116]. Also, in diabetes related with ED, the overexpression of RhoA/Rho-kinase signaling was observed in penile tissue from diabetic rats, and ROCK1 protein expression is increased but not ROCK2. In this study, the administration of testosterone in diabetic rats was able to treat hypogonadism and maintain erectile function by

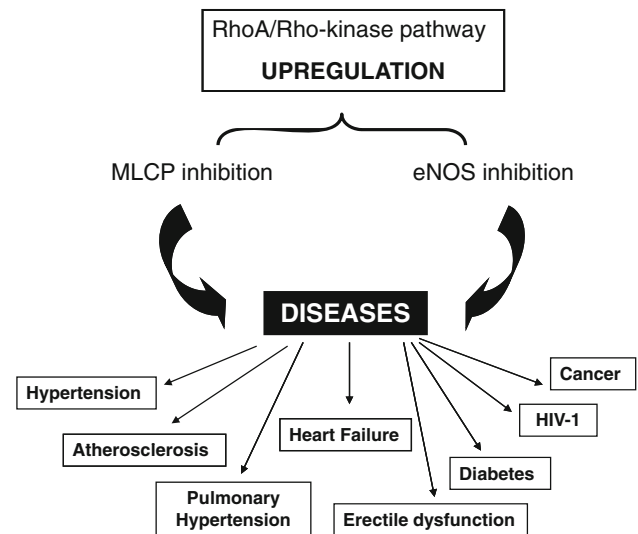


normalizing RhoA/Rho-kinase pathway upregulation [165]. Nevertheless, the role of androgens on the vasoconstrictor action of components of the RhoA/Rho-kinase pathway involved in the erectile process remains to be elucidated.

Since it was demonstrated that Y-27632 improved erectile function in aged rats, it has been suggested that the RhoA/Rho-kinase pathway could be important in the pathology of ED associated with age [166]. However, it is difficult to know exactly the role of RhoA/Rho-kinase in ED because this inhibitor has effects on multiple kinases involved in smooth muscle contraction. Nevertheless, a study used adeno-associated viral gene transfer of dominant-negative RhoA mutant (T19NRhoA) into the rat corpus cavernosum, as a tool to target specifically RhoA. The results indicated increased intracavernosal blood pressure (ICP), suggesting that the ED-associated age process involves increased RhoA/Rho-kinase pathway signaling [167]. The activation of neuronal nitric oxide synthase (nNOS) in corpus cavernosum tissue decreased with ageing. Additionally, it has been suggested that, in old rats, the imbalance between nNOS activity and Rho-kinase could be associated with impaired erectile function [11]. Taken together, it is clear that RhoA/Rho-kinase activity is a fundamental component to keep the penis in the non-erect state, and this pathway is upregulated in ED. Also, the essential balance between contraction and relaxation in the penis, which is maintained by the RhoA/Rho-kinase and NO/cyclicGMP pathways, is modified in this pathology.

## Conclusion

Due to the rapidly growing number of studies implicating the RhoA/Rho-kinase pathway in various pathologies, it is undeniable that Rho-kinase is an important therapeutic target. Specific and potent pharmacological modulators of various steps of the RhoA/Rho-kinase signaling pathway are critically needed for treatment intervention in cardiovascular diseases, neurological disorders, and cancer progression. However, a greater understanding of the physiological role of each of the Rho-kinase isoforms and the development of isoform-specific inhibitors are needed to achieve these goals. Taken together, the connection between many diseases and RhoA/Rho-kinase seems to be in the upregulation of this pathway. Increased activation of MLCK is a consequence of RhoA/Rho-kinase pathway upregulation leading to MLCP and eNOS inhibition (Fig. 2). Both events facilitate vascular contraction, along with interfering in other cellular responses, such as focal adhesion, migration, gene expression, and apoptosis signaling, which contribute to the development of several pathologies (Fig. 2) [2, 4, 44, 72, 168]. Nevertheless, many



**Fig. 2** Upregulation of RhoA/Rho-kinase pathway is a key link among many diseases. MLCP and eNOS activity are targeted by RhoA/Rho-kinase upregulation, leading to alterations in many physiological processes associated with these two molecules, resulting in various disease states

questions about how upregulation of RhoA/Rho-kinase interferes in the each pathology remain unclear.

## References

1. Amano M, Fukata Y, Kaibuchi K (2000) Regulation and functions of Rho-associated kinase. *Exp Cell Res* 261:44–51
2. Ridley AJ (2001) Rho family proteins: coordinating cell responses. *Trends Cell Biol* 11:471–477
3. Nakagawa O, Fujisawa K, Ishizaki T, Saito Y, Nakao K, Narumiya S (1996) ROCK-I and ROCK-II, two isoforms of Rho-associated coiled-coil forming protein serine/threonine kinase in mice. *FEBS Lett* 392:189–193
4. Wettschreck N, Offermanns S (2002) Rho/Rho-kinase mediated signaling in physiology and pathophysiology. *J Mol Med* 80:629–638
5. Shimokawa H (2002) Rho-kinase as a novel therapeutic target in treatment of cardiovascular diseases. *J Cardiovasc Pharmacol* 39:319–327
6. Somlyo AP, Somlyo AV (2004) Signal transduction through the RhoA/Rho-kinase pathway in smooth muscle. *J Muscle Res Cell Motil* 25:613–615
7. Kureishi Y, Kobayashi S, Amano M, Kimura K, Kanaide H, Nakano T, Kaibuchi K, Ito M (1997) Rho-associated kinase directly induces smooth muscle contraction through myosin light chain phosphorylation. *J Biol Chem* 272:12257–12260
8. Berridge MJ (2008) Smooth muscle cell calcium activation mechanisms. *J Physiol* 586:5047–5061
9. Hiroki J, Shimokawa H, Higashi M, Morikawa K, Kandabashi T, Kawamura N, Kubota T, Ichiki T, Amano M, Kaibuchi K, Takeshita A (2004) Inflammatory stimuli upregulate Rho-kinase in human coronary vascular smooth muscle cells. *J Mol Cell Cardiol* 37:537–546
10. Barman SA, Zhu S, White RE (2009) RhoA/Rho-kinase signaling: a therapeutic target in pulmonary hypertension. *Vasc Health Risk Manag* 5:663–671

11. Gao BH, Zhao ST, Meng FW, Shi BK, Liu YQ, Xu ZS (2007) Y-27632 improves the erectile dysfunction with ageing in SD rats through adjusting the imbalance between nNo and the Rho-kinase pathways. *Andrologia* 39:146–150
12. Nakamura Y, Kaneto H, Miyatsuka T, Matsuoka TA, Matsuoka M, Node K, Hori M, Yamasaki Y (2006) Marked increase of insulin gene transcription by suppression of the Rho/Rho-kinase pathway. *Biochem Biophys Res Commun* 350:68–73
13. Chang YW, Bean RR, Jakobi R (2009) Targeting RhoA/Rho kinase and p21-activated kinase signaling to prevent cancer development and progression. *Recent Pat Anticancer Drug Discov* 4:110–124
14. LoGrasso PV, Feng Y (2009) Rho kinase (ROCK) inhibitors and their application to inflammatory disorders. *Curr Top Med Chem* 9:704–723
15. Liao JK, Seto M, Noma K (2007) Rho kinase (ROCK) inhibitors. *J Cardiovasc Pharmacol* 50:17–24
16. Sorokina EM, Chernoff J (2005) Rho-GTPases: new members, new pathways. *J Cell Biochem* 94:225–231
17. Fukata Y, Amano M, Kaibuchi K (2001) Rho-Rho-kinase pathway in smooth muscle contraction and cytoskeletal reorganization of non-muscle cells. *Trends Pharmacol Sci* 22:32–39
18. Jaffe AB, Hall A (2005) Rho GTPases: biochemistry and biology. *Annu Rev Cell Dev Biol* 21:247–269
19. Riento K, Ridley AJ (2003) Rocks: multifunctional kinases in cell behaviour. *Nat Rev Mol Cell Biol* 4:446–456
20. Komander D, Garg R, Wan PT, Ridley AJ, Barford D (2008) Mechanism of multi-site phosphorylation from a ROCK-I: RhoE complex structure. *EMBO J* 27:3175–3185
21. Riento K, Totty N, Villalonga P, Garg R, Guasch R, Ridley AJ (2005) RhoE function is regulated by ROCK I-mediated phosphorylation. *EMBO J* 24:1170–1180
22. Kurokawa T, Yumiya Y, Fujisawa H, Shirao S, Kashiwagi S, Sato M, Kishi H, Miwa S, Mogami K, Kato S, Akimura T, Soma M, Ogasawara K, Ogawa A, Kobayashi S, Suzuki M (2009) Elevated concentrations of sphingosylphosphorylcholine in cerebrospinal fluid after subarachnoid hemorrhage: a possible role as a spasmogen. *J Clin Neurosci* 16:1064–1068
23. Matsui T, Amano M, Yamamoto T, Chihara K, Nakafuku M, Ito M, Nakano T, Okawa K, Iwamatsu A, Kaibuchi K (1996) Rho-associated kinase, a novel serine/threonine kinase, as a putative target for small GTP binding protein Rho. *EMBO J* 15:2208–2216
24. Yoneda A, Multhaupt HA, Couchman JR (2005) The Rho kinases I and II regulate different aspects of myosin II activity. *J Cell Biol* 170:443–453
25. Sebbagh M, Renvoize C, Hamelin J, Riche N, Bertoglio J, Breard J (2001) Caspase-3-mediated cleavage of ROCK I induces MLC phosphorylation and apoptotic membrane blebbing. *Nat Cell Biol* 3:346–352
26. Wang Y, Zheng XR, Riddick N, Bryden M, Baur W, Zhang X, Surks HK (2009) ROCK isoform regulation of myosin phosphatase and contractility in vascular smooth muscle cells. *Circ Res* 104:531–540
27. Thumkeo D, Keel J, Ishizaki T, Hirose M, Nonomura K, Oshima H, Oshima M, Taketo MM, Narumiya S (2003) Targeted disruption of the mouse rho-associated kinase 2 gene results in intrauterine growth retardation and fetal death. *Mol Cell Biol* 23:5043–5055
28. Loirand G, Guerin P, Pacaud P (2006) Rho kinases in cardiovascular physiology and pathophysiology. *Circ Res* 98:322–334
29. Wei L, Roberts W, Wang L, Yamada M, Zhang S, Zhao Z, Rivkees SA, Schwartz RJ, Imanaka-Yoshida K (2001) Rho kinases play an obligatory role in vertebrate embryonic organogenesis. *Development* 128:2953–2962
30. Di Cunto F, Imarisio S, Hirsch E, Broccoli V, Bulfone A, Migheli A, Atzori C, Turco E, Triolo R, Dotto GP, Silengo L, Altruda F (2000) Defective neurogenesis in citron kinase knockout mice by altered cytokinesis and massive apoptosis. *Neuron* 28:115–127
31. Yamashita K, Kotani Y, Nakajima Y, Shimazawa M, Yoshimura S, Nakashima S, Iwama T, Hara H (2007) Fasudil, a Rho kinase (ROCK) inhibitor, protects against ischemic neuronal damage in vitro and in vivo by acting directly on neurons. *Brain Res* 1154:215–224
32. Leung T, Manser E, Tan L, Lim L (1995) A novel serine/threonine kinase binding the Ras-related RhoA GTPase which translocates the kinase to peripheral membranes. *J Biol Chem* 270:29051–29054
33. Chevrier V, Piel M, Collomb N, Saoudi Y, Frank R, Paintrand M, Narumiya S, Bornens M, Job D (2002) The Rho-associated protein kinase p160ROCK is required for centrosome positioning. *J Cell Biol* 157:807–817
34. Sah VP, Seasholtz TM, Sagi SA, Brown JH (2000) The role of Rho in G protein-coupled receptor signal transduction. *Annu Rev Pharmacol Toxicol* 40:459–489
35. Mizuno Y, Isotani E, Huang J, Ding H, Stull JT, Kamm KE (2008) Myosin light chain kinase activation and calcium sensitization in smooth muscle in vivo. *Am J Physiol Cell Physiol* 295:C358–C364
36. Newton AC (1995) Protein kinase C: structure, function, and regulation. *J Biol Chem* 270:28495–28498
37. Hilgers RH, Webb RC (2005) Molecular aspects of arterial smooth muscle contraction: focus on Rho. *Exp Biol Med* (Maywood) 230:829–835
38. Muranyi A, Derkach D, Erdodi F, Kiss A, Ito M, Hartshorne DJ (2005) Phosphorylation of Thr695 and Thr850 on the myosin phosphatase target subunit: inhibitory effects and occurrence in A7r5 cells. *FEBS Lett* 579:6611–6615
39. Eto M, Ohmori T, Suzuki M, Furuya K, Morita F (1995) A novel protein phosphatase-I inhibitory protein potentiated by protein kinase C. Isolation from porcine aorta media and characterization. *J Biochem* 118:1104–1107
40. Koyama M, Ito M, Feng J, Seko T, Shiraki K, Takase K, Hartshorne DJ, Nakano T (2000) Phosphorylation of CPI-17, an inhibitory phosphoprotein of smooth muscle myosin phosphatase, by Rho-kinase. *FEBS Lett* 475:197–200
41. Surks HK, Riddick N, Ohtani K (2005) M-RIP targets myosin phosphatase to stress fibers to regulate myosin light chain phosphorylation in vascular smooth muscle cells. *J Biol Chem* 280:42543–42551
42. Koga Y, Ikebe M (2005) p116Rip decreases myosin II phosphorylation by activating myosin light chain phosphatase and by inactivating RhoA. *J Biol Chem* 280:4983–4991
43. Jin L, Ying Z, Hilgers RH, Yin J, Zhao X, Imig JD, Webb RC (2006) Increased RhoA/Rho-kinase signaling mediates spontaneous tone in aorta from angiotensin II-induced hypertensive rats. *J Pharmacol Exp Ther* 318:288–295
44. Zhou Q, Liao JK (2009) Rho kinase: an important mediator of atherosclerosis and vascular disease. *Curr Pharm Des* 15:3108–3115
45. Hamid SA, Bower HS, Baxter GF (2007) Rho kinase activation plays a major role as a mediator of irreversible injury in reperfused myocardium. *Am J Physiol Heart Circ Physiol* 292:H2598–H2606
46. Rikitake Y, Kim HH, Huang Z, Seto M, Yano K, Asano T, Moskowitz MA, Liao JK (2005) Inhibition of Rho kinase (ROCK) leads to increased cerebral blood flow and stroke protection. *Stroke* 36:2251–2257
47. Sato M, Tani E, Fujikawa H, Kaibuchi K (2000) Involvement of Rho-kinase-mediated phosphorylation of myosin light chain in enhancement of cerebral vasospasm. *Circ Res* 87:195–200

48. Higashi M, Shimokawa H, Hattori T, Hiroki J, Mukai Y, Morikawa K, Ichiki T, Takahashi S, Takeshita A (2003) Long-term inhibition of Rho-kinase suppresses angiotensin II-induced cardiovascular hypertrophy in rats in vivo: effect on endothelial NAD(P)H oxidase system. *Circ Res* 93:767–775
49. Bao W, Hu E, Tao L, Boyce R, Mirabile R, Thudium DT, Ma XL, Willette RN, Yue TL (2004) Inhibition of Rho-kinase protects the heart against ischemia/reperfusion injury. *Cardiovasc Res* 61:548–558
50. Miyata K, Shimokawa H, Kandabashi T, Higo T, Morishige K, Eto Y, Egashira K, Kaibuchi K, Takeshita A (2000) Rho-kinase is involved in macrophage-mediated formation of coronary vascular lesions in pigs in vivo. *Arterioscler Thromb Vasc Biol* 20:2351–2358
51. Ming XF, Viswambharan H, Barandier C, Ruffieux J, Kaibuchi K, Rusconi S, Yang Z (2002) Rho GTPase/Rho kinase negatively regulates endothelial nitric oxide synthase phosphorylation through the inhibition of protein kinase B/Akt in human endothelial cells. *Mol Cell Biol* 22:8467–8477
52. Wolfrum S, Dendorfer A, Rikitake Y, Stalker TJ, Gong Y, Scalia R, Dominiak P, Liao JK (2004) Inhibition of Rho-kinase leads to rapid activation of phosphatidylinositol 3-kinase/protein kinase Akt and cardiovascular protection. *Arterioscler Thromb Vasc Biol* 24:1842–1847
53. Dimmeler S, Fleming I, Fisslthaler B, Hermann C, Busse R, Zeiher AM (1999) Activation of nitric oxide synthase in endothelial cells by Akt-dependent phosphorylation. *Nature* 399:601–605
54. Chen ZP, Mitchelhill KI, Michell BJ, Stapleton D, Rodriguez-Crespo I, Witters LA, Power DA, Ortiz de Montellano PR, Kemp BE (1999) AMP-activated protein kinase phosphorylation of endothelial NO synthase. *FEBS Lett* 443:285–289
55. Kishi T, Hirooka Y, Masumoto A, Ito K, Kimura Y, Inokuchi K, Tagawa T, Shimokawa H, Takeshita A, Sunagawa K (2005) Rho-kinase inhibitor improves increased vascular resistance and impaired vasodilation of the forearm in patients with heart failure. *Circulation* 111:2741–2747
56. Shimokawa H, Hiramori K, Inuma H, Hosoda S, Kishida H, Osada H, Katagiri T, Yamauchi K, Yui Y, Minamino T, Nakashima M, Kato K (2002) Anti-anginal effect of fasudil, a Rho-kinase inhibitor, in patients with stable effort angina: a multicenter study. *J Cardiovasc Pharmacol* 40:751–761
57. Miao L, Calvert JW, Tang J, Parent AD, Zhang JH (2001) Age-related RhoA expression in blood vessels of rats. *Mech Ageing Dev* 122:1757–1770
58. Kai H, Kudo H, Takayama N, Yasuoka S, Kajimoto H, Imaizumi T (2009) Large blood pressure variability and hypertensive cardiac remodeling—role of cardiac inflammation. *Circ J* 73:2198–2203
59. Cousin M, Custaud MA, Baron-Menguy C, Toutain B, Dumont O, Guihot AL, Vessieres E, Subra JF, Henrion D, Loufrani L (2010) Role of angiotensin II in the remodeling induced by a chronic increase in flow in rat mesenteric resistance arteries. *Hypertension* 55:109–115
60. Wesselman JP, De Mey JG (2002) Angiotensin and cytoskeletal proteins: role in vascular remodeling. *Curr Hypertens Rep* 4:63–70
61. Wynne BM, Chiao C-W, Webb RC (2009) Vascular smooth muscle cell signaling mechanism for construction to angiotensin II and endothelin-1. *J Am Soc Hypertension* 3:84–95
62. Zhong Y, Hennig B, Toborek M (2010) Intact lipid rafts regulate HIV-1 Tat protein-induced activation of the Rho signaling and upregulation of P-glycoprotein in brain endothelial cells. *J Cereb Blood Flow Metab* 30:522–533
63. Xiao H, Neuveut C, Tiffany HL, Benkirane M, Rich EA, Murphy PM, Jeang KT (2000) Selective CXCR4 antagonism by Tat: implications for in vivo expansion of coreceptor use by HIV-1. *Proc Natl Acad Sci USA* 97:11466–11471
64. Fukumoto Y, Tawara S, Shimokawa H (2007) Recent progress in the treatment of pulmonary arterial hypertension: expectation for rho-kinase inhibitors. *Tohoku J Exp Med* 211:309–320
65. Wakino S, Kanda T, Hayashi K (2005) Rho/Rho kinase as a potential target for the treatment of renal disease. *Drug News Perspect* 18:639–643
66. Chobanian AV, Bakris GL, Black HR, Cushman WC, Green LA, Izzo JL Jr, Jones DW, Materson BJ, Oparil S, Wright JT Jr, Roccella EJ (2003) The seventh report of the joint national committee on prevention, detection, evaluation, and treatment of high blood pressure: the JNC 7 report. *JAMA* 289:2560–2572
67. Kokubo Y, Kamide K (2009) High-normal blood pressure and the risk of cardiovascular disease. *Circ J* 73:1381–1385
68. Masumoto A, Hirooka Y, Shimokawa H, Hironaga K, Setoguchi S, Takeshita A (2001) Possible involvement of Rho-kinase in the pathogenesis of hypertension in humans. *Hypertension* 38:1307–1310
69. Uehata M, Ishizaki T, Satoh H, Ono T, Kawahara T, Morishita T, Tamakawa H, Yamagami K, Inui J, Maekawa M, Narumiya S (1997) Calcium sensitization of smooth muscle mediated by a Rho-associated protein kinase in hypertension. *Nature* 389:990–994
70. Ito K, Hirooka Y, Sakai K, Kishi T, Kaibuchi K, Shimokawa H, Takeshita A (2003) Rho/Rho-kinase pathway in brain stem contributes to blood pressure regulation via sympathetic nervous system: possible involvement in neural mechanisms of hypertension. *Circ Res* 92:1337–1343
71. Ito K, Hirooka Y, Kishi T, Kimura Y, Kaibuchi K, Shimokawa H, Takeshita A (2004) Rho/Rho-kinase pathway in the brain-stem contributes to hypertension caused by chronic nitric oxide synthase inhibition. *Hypertension* 43:156–162
72. Takemoto M, Sun J, Hiroki J, Shimokawa H, Liao JK (2002) Rho-kinase mediates hypoxia-induced downregulation of endothelial nitric oxide synthase. *Circulation* 106:57–62
73. Shin HK, Salomone S, Potts EM, Lee SW, Millican E, Noma K, Huang PL, Boas DA, Liao JK, Moskowitz MA, Ayata C (2007) Rho-kinase inhibition acutely augments blood flow in focal cerebral ischemia via endothelial mechanisms. *J Cereb Blood Flow Metab* 27:998–1009
74. Mukai Y, Shimokawa H, Matoba T, Kandabashi T, Satoh S, Hiroki J, Kaibuchi K, Takeshita A (2001) Involvement of Rho-kinase in hypertensive vascular disease: a novel therapeutic target in hypertension. *FASEB J* 15:1062–1064
75. Moriki N, Ito M, Seko T, Kureishi Y, Okamoto R, Nakakuki T, Kongo M, Isaka N, Kaibuchi K, Nakano T (2004) RhoA activation in vascular smooth muscle cells from stroke-prone spontaneously hypertensive rats. *Hypertens Res* 27:263–270
76. Hahmann C, Schroeter T (2010) Rho-kinase inhibitors as therapeutics: from pan inhibition to isoform selectivity. *Cell Mol Life Sci* 67:171–177
77. Lohn M, Plettenburg O, Ivashchenko Y, Kannt A, Hofmeister A, Kadereit D, Schaefer M, Linz W, Kohlmann M, Herbert JM, Janiak P, O'Connor SE, Ruetten H (2009) Pharmacological characterization of SAR407899, a novel rho-kinase inhibitor. *Hypertension* 54:676–683
78. Mallat Z, Gojova A, Sauzeau V, Brun V, Silvestre JS, Esposito B, Merval R, Groux H, Loirand G, Tedgui A (2003) Rho-associated protein kinase contributes to early atherosclerotic lesion formation in mice. *Circ Res* 93:884–888
79. Wang HW, Liu PY, Oyama N, Rikitake Y, Kitamoto S, Gitlin J, Liao JK, Boisvert WA (2008) Deficiency of ROCK1 in bone marrow-derived cells protects against atherosclerosis in LDLR<sup>-/-</sup> mice. *FASEB J* 22:3561–3570

80. Cannon CP, Braunwald E, McCabe CH, Rader DJ, Rouleau JL, Belder R, Joyal SV, Hill KA, Pfeffer MA, Skene AM (2004) Intensive versus moderate lipid lowering with statins after acute coronary syndromes. *N Engl J Med* 350:1495–1504
81. Ito T, Ikeda U, Shimpo M, Ohki R, Takahashi M, Yamamoto K, Shimada K (2002) HMG-CoA reductase inhibitors reduce interleukin-6 synthesis in human vascular smooth muscle cells. *Cardiovasc Drugs Ther* 16:121–126
82. Rikitake Y, Liao JK (2005) Rho-kinase mediates hyperglycemia-induced plasminogen activator inhibitor-1 expression in vascular endothelial cells. *Circulation* 111:3261–3268
83. Laufs U, Endres M, Stagliano N, Amin-Hanjani S, Chui DS, Yang SX, Simoncini T, Yamada M, Rabkin E, Allen PG, Huang PL, Bohm M, Schoen FJ, Moskowitz MA, Liao JK (2000) Neuroprotection mediated by changes in the endothelial actin cytoskeleton. *J Clin Invest* 106:15–24
84. Nohria A, Prsic A, Liu PY, Okamoto R, Creager MA, Selwyn A, Liao JK, Ganz P (2009) Statins inhibit Rho kinase activity in patients with atherosclerosis. *Atherosclerosis* 205:517–521
85. Reikhter M, Chandrasekhar K, Gifford-Moore D, Huang XD, Rutherford P, Hanson J, Kauffman R (2007) Immunohistochemical analysis of target proteins of Rho-kinase in a mouse model of accelerated atherosclerosis. *Exp Clin Cardiol* 12:169–174
86. Mueller BK, Mack H, Teusch N (2005) Rho kinase, a promising drug target for neurological disorders. *Nat Rev Drug Discov* 4:387–398
87. Borisoff JF, Chan CC, Hiebert GW, Oschepok L, Robertson GS, Zamboni R, Steeves JD, Tetzlaff W (2003) Suppression of Rho-kinase activity promotes axonal growth on inhibitory CNS substrates. *Mol Cell Neurosci* 22:405–416
88. Masumoto A, Mohri M, Shimokawa H, Urakami L, Usui M, Takeshita A (2002) Suppression of coronary artery spasm by the Rho-kinase inhibitor fasudil in patients with vasospastic angina. *Circulation* 105:1545–1547
89. Mills JC, Stone NL, Pittman RN (1999) Extranuclear apoptosis. The role of the cytoplasm in the execution phase. *J Cell Biol* 146:703–708
90. Satoh S, Toshima Y, Hitomi A, Ikegaki I, Seto M, Asano T (2008) Wide therapeutic time window for Rho-kinase inhibition therapy in ischemic brain damage in a rat cerebral thrombosis model. *Brain Res* 1193:102–108
91. Hitomi A, Satoh S, Ikegaki I, Suzuki Y, Shibuya M, Asano T (2000) Hemorheological abnormalities in experimental cerebral ischemia and effects of protein kinase inhibitor on blood fluidity. *Life Sci* 67:1929–1939
92. Satoh S, Kobayashi T, Hitomi A, Ikegaki I, Suzuki Y, Shibuya M, Yoshida J, Asano T (1999) Inhibition of neutrophil migration by a protein kinase inhibitor for the treatment of ischemic brain infarction. *Jpn J Pharmacol* 80:41–48
93. Ding J, Li QY, Yu JZ, Wang X, Sun CH, Lu CZ, Xiao BG (2010) Fasudil, a Rho kinase inhibitor, drives mobilization of adult neural stem cells after hypoxia/reoxygenation injury in mice. *Mol Cell Neurosci* 43:201–208
94. Suematsu N, Satoh S, Kinugawa S, Tsutsui H, Hayashidani S, Nakamura R, Egashira K, Makino N, Takeshita A (2001) Alpha1-adrenoceptor-Gq-RhoA signaling is upregulated to increase myofibrillar  $Ca^{2+}$  sensitivity in failing hearts. *Am J Physiol Heart Circ Physiol* 281:H637–H646
95. Kobayashi N, Horinaka S, Mita S, Nakano S, Honda T, Yoshida K, Kobayashi T, Matsuoka H (2002) Critical role of Rho-kinase pathway for cardiac performance and remodeling in failing rat hearts. *Cardiovasc Res* 55:757–767
96. Demiryurek S, Kara AF, Celik A, Babul A, Tarakcioglu M, Demiryurek AT (2005) Effects of fasudil, a Rho-kinase inhibitor, on myocardial preconditioning in anesthetized rats. *Eur J Pharmacol* 527:129–140
97. Sanada S, Asanuma H, Tsukamoto O, Minamino T, Node K, Takashima S, Fukushima T, Ogai A, Shinozaki Y, Fujita M, Hirata A, Okuda H, Shimokawa H, Tomoike H, Hori M, Kitakaze M (2004) Protein kinase A as another mediator of ischemic preconditioning independent of protein kinase C. *Circulation* 110:51–57
98. Manintveld OC, Verdouw PD, Duncker DJ (2007) The RISK of ROCK. *Am J Physiol Heart Circ Physiol* 292:H2563–H2565
99. Kobayashi N, Takeshima H, Fukushima H, Koguchi W, Mamada Y, Hirata H, Machida Y, Shinoda M, Suzuki N, Yokotsuka F, Tabei K, Matsuoka H (2009) Cardioprotective effects of pitavastatin on cardiac performance and remodeling in failing rat hearts. *Am J Hypertension* 22:176–182
100. Dulak J, Jozkowicz A (2005) Anti-angiogenic and anti-inflammatory effects of statins: relevance to anti-cancer therapy. *Curr Cancer Drug Targets* 5:579–594
101. Fritz G, Kaina B (2006) Rho GTPases: promising cellular targets for novel anticancer drugs. *Curr Cancer Drug Targets* 6:1–14
102. Isezu SA (2006) The metabolic syndrome: review of current concepts. *Niger Postgrad Med J* 13:247–255
103. Okon EB, Szado T, Laher I, McManus B, van Breemen C (2003) Augmented contractile response of vascular smooth muscle in a diabetic mouse model. *J Vasc Res* 40:520–530
104. Sowers JR, Lester MA (1999) Diabetes and cardiovascular disease. *Diabetes Care* 22(Suppl 3):C14–C20
105. Goldin A, Beckman JA, Schmidt AM, Creager MA (2006) Advanced glycation end products: sparking the development of diabetic vascular injury. *Circulation* 114:597–605
106. Hirose A, Tanikawa T, Mori H, Okada Y, Tanaka Y (2010) Advanced glycation end products increase endothelial permeability through the RAGE/Rho signaling pathway. *FEBS Lett* 584:61–66
107. Massey AR, Miao L, Smith BN, Liu J, Kusaka I, Zhang JH, Tang J (2003) Increased RhoA translocation in renal cortex of diabetic rats. *Life Sci* 72:2943–2952
108. Peng F, Wu D, Gao B, Ingram AJ, Zhang B, Chorneyko K, McKenzie R, Krepinsky JC (2008) RhoA/Rho-kinase contribute to the pathogenesis of diabetic renal disease. *Diabetes* 57:1683–1692
109. Nuno DW, Harrod JS, Lamping KG (2009) Sex-dependent differences in Rho activation contribute to contractile dysfunction in type 2 diabetic mice. *Am J Physiol Heart Circ Physiol* 297:H1469–H1477
110. Kanda T, Wakino S, Homma K, Yoshioka K, Tatamatsu S, Hasegawa K, Takamatsu I, Sugano N, Hayashi K, Saruta T (2006) Rho-kinase as a molecular target for insulin resistance and hypertension. *FASEB J* 20:169–171
111. Hu E, Lee D (2005) Rho kinase as potential therapeutic target for cardiovascular diseases: opportunities and challenges. *Expert Opin Ther Targets* 9:715–736
112. Kolavennu V, Zeng L, Peng H, Wang Y, Danesh FR (2008) Targeting of RhoA/ROCK signaling ameliorates progression of diabetic nephropathy independent of glucose control. *Diabetes* 57:714–723
113. Furukawa N, Ongusaha P, Jahng WJ, Araki K, Choi CS, Kim HJ, Lee YH, Kaibuchi K, Kahn BB, Masuzaki H, Kim JK, Lee SW, Kim YB (2005) Role of Rho-kinase in regulation of insulin action and glucose homeostasis. *Cell Metab* 2:119–129
114. Lee DH, Shi J, Jeoung NH, Kim MS, Zabolotny JM, Lee SW, White MF, Wei L, Kim YB (2009) Targeted disruption of ROCK1 causes insulin resistance in vivo. *J Biol Chem* 284:11776–11780
115. Chang S, Hypolite JA, Changolkar A, Wein AJ, Chacko S, DiSanto ME (2003) Increased contractility of diabetic rabbit corpora smooth muscle in response to endothelin is mediated via Rho-kinase beta. *Int J Impot Res* 15:53–62



116. Bivalacqua TJ, Champion HC, Usta MF, Celtek S, Chitale K, Webb RC, Lewis RL, Mills TM, Hellstrom WJ, Kadowitz PJ (2004) RhoA/Rho-kinase suppresses endothelial nitric oxide synthase in the penis: a mechanism for diabetes-associated erectile dysfunction. *Proc Natl Acad Sci USA* 101:9121–9126
117. Sahai E, Marshall CJ (2002) RHO-GTPases and cancer. *Nat Rev Cancer* 2:133–142
118. Keely PJ (2001) Rho GTPases as early markers for tumour progression. *Lancet* 358:1744–1745
119. Jaffe AB, Hall A (2002) Rho GTPases in transformation and metastasis. *Adv Cancer Res* 84:57–80
120. Abraham MT, Kuriakose MA, Sacks PG, Yee H, Chiriboga L, Bearer EL, Delacure MD (2001) Motility-related proteins as markers for head and neck squamous cell cancer. *Laryngoscope* 111:1285–1289
121. Itoh K, Yoshioka K, Akedo H, Uehata M, Ishizaki T, Narumiya S (1999) An essential part for Rho-associated kinase in the transcellular invasion of tumor cells. *Nat Med* 5:221–225
122. Ogata S, Morishige K, Sawada K, Hashimoto K, Mabuchi S, Kawase C, Ooyagi C, Sakata M, Kimura T (2009) Fasudil inhibits lysophosphatidic acid-induced invasiveness of human ovarian cancer cells. *Int J Gynecol Cancer* 19:1473–1480
123. Somlyo AV, Bradshaw D, Ramos S, Murphy C, Myers CE, Somlyo AP (2000) Rho-kinase inhibitor retards migration and in vivo dissemination of human prostate cancer cells. *Biochem Biophys Res Commun* 269:652–659
124. Lin MT, Lin BR, Chang CC, Chu CY, Su HJ, Chen ST, Jeng YM, Kuo ML (2007) IL-6 induces AGS gastric cancer cell invasion via activation of the c-Src/RhoA/ROCK signaling pathway. *Int J Cancer* 120:2600–2608
125. Yoshioka K, Matsumura F, Akedo H, Itoh K (1998) Small GTP-binding protein Rho stimulates the actomyosin system, leading to invasion of tumor cells. *J Biol Chem* 273:5146–5154
126. del Peso L, Hernandez-Alcoceba R, Embade N, Carnero A, Esteve P, Paje C, Lacal JC (1997) Rho proteins induce metastatic properties in vivo. *Oncogene* 15:3047–3057
127. Sun HW, Tong SL, He J, Wang Q, Zou L, Ma SJ, Tan HY, Luo JF, Wu HX (2007) RhoA and RhoC-siRNA inhibit the proliferation and invasiveness activity of human gastric carcinoma by Rho/PI3K/Akt pathway. *World J Gastroenterol* 13:3517–3522
128. Clark EA, Golub TR, Lander ES, Hynes RO (2000) Genomic analysis of metastasis reveals an essential role for RhoC. *Nature* 406:532–535
129. Coleman ML, Olson MF (2002) Rho GTPase signalling pathways in the morphological changes associated with apoptosis. *Cell Death Differ* 9:493–504
130. Vishnubhotla R, Sun S, Huq J, Bulic M, Ramesh A, Guzman G, Cho M, Glover SC (2007) ROCK-II mediates colon cancer invasion via regulation of MMP-2 and MMP-13 at the site of invadopodia as revealed by multiphoton imaging. *Lab Invest* 87:1149–1158
131. Zhou Z, Meng Y, Asrar S, Todorovski Z, Jia Z (2009) A critical role of Rho-kinase ROCK2 in the regulation of spine and synaptic function. *Neuropharmacology* 56:81–89
132. Sterpetti P, Marucci L, Candelaesi C, Toksoz D, Alpini G, Ugili L, Baroni GS, Macarri G, Benedetti A (2006) Cell proliferation and drug resistance in hepatocellular carcinoma are modulated by Rho GTPase signals. *Am J Physiol Gastrointest Liver Physiol* 290:G624–G632
133. Lu Q, Longo FM, Zhou H, Massa SM, Chen YH (2009) Signaling through Rho GTPase pathway as viable drug target. *Curr Med Chem* 16:1355–1365
134. Fukumoto Y, Matoba T, Ito A, Tanaka H, Kishi T, Hayashidani S, Abe K, Takeshita A, Shimokawa H (2005) Acute vasodilator effects of a Rho-kinase inhibitor, fasudil, in patients with severe pulmonary hypertension. *Heart* 91:391–392
135. Li F, Xia W, Li A, Zhao C, Sun R (2007) Long-term inhibition of Rho kinase with fasudil attenuates high flow induced pulmonary artery remodeling in rats. *Pharmacol Res* 55:64–71
136. Barst RJ, McGoon M, Torbicki A, Sitbon O, Krowka MJ, Olschewski H, Gaine S (2004) Diagnosis and differential assessment of pulmonary arterial hypertension. *J Am Coll Cardiol* 43:40S–47S
137. Oka M, Homma N, Taraseviciene-Stewart L, Morris KG, Kraskauskas D, Burns N, Voelkel NF, McMurtry IF (2007) Rho kinase-mediated vasoconstriction is important in severe occlusive pulmonary arterial hypertension in rats. *Circ Res* 100:923–929
138. Do e Z, Fukumoto Y, Takaki A, Tawara S, Ohashi J, Nakano M, Tada T, Saji K, Sugimura K, Fujita H, Hoshikawa Y, Nawata J, Kondo T, Shimokawa H (2009) Evidence for Rho-kinase activation in patients with pulmonary arterial hypertension. *Circ J* 73:1731–1739
139. Li F, Xia W, Yuan S, Sun R (2009) Acute inhibition of Rho-kinase attenuates pulmonary hypertension in patients with congenital heart disease. *Pediatr Cardiol* 30:363–366
140. Fujita H, Fukumoto Y, Saji K, Sugimura K, Demachi J, Nawata J, Shimokawa H (2010) Acute vasodilator effects of inhaled fasudil, a specific Rho-kinase inhibitor, in patients with pulmonary arterial hypertension. *Heart Vessel* 25:144–149
141. Guilluy C, Sauzeau V, Rolli-Derkinderen M, Guerin P, Sagan C, Pacaud P, Loirand G (2005) Inhibition of RhoA/Rho kinase pathway is involved in the beneficial effect of sildenafil on pulmonary hypertension. *Br J Pharmacol* 146:1010–1018
142. Hyvelin JM, Howell K, Nichol A, Costello CM, Preston RJ, McLoughlin P (2005) Inhibition of Rho-kinase attenuates hypoxia-induced angiogenesis in the pulmonary circulation. *Circ Res* 97:185–191
143. Nagaoka T, Fagan KA, Gebb SA, Morris KG, Suzuki T, Shimokawa H, McMurtry IF, Oka M (2005) Inhaled Rho kinase inhibitors are potent and selective vasodilators in rat pulmonary hypertension. *Am J Respir Crit Care Med* 171:494–499
144. Homma N, Nagaoka T, Karoor V, Imamura M, Taraseviciene-Stewart L, Walker LA, Fagan KA, McMurtry IF, Oka M (2008) Involvement of RhoA/Rho kinase signaling in protection against monocrotaline-induced pulmonary hypertension in pneumonectomized rats by dehydroepiandrosterone. *Am J Physiol Lung Cell Mol Physiol* 295:L71–L78
145. Nagaoka T, Gebb SA, Karoor V, Homma N, Morris KG, McMurtry IF, Oka M (2006) Involvement of RhoA/Rho kinase signaling in pulmonary hypertension of the fawn-hooded rat. *J Appl Physiol* 100:996–1002
146. Broughton BR, Walker BR, Resta TC (2008) Chronic hypoxia induces Rho kinase-dependent myogenic tone in small pulmonary arteries. *Am J Physiol Lung Cell Mol Physiol* 294:L797–L806
147. Fagan KA, Oka M, Bauer NR, Gebb SA, Ivy DD, Morris KG, McMurtry IF (2004) Attenuation of acute hypoxic pulmonary vasoconstriction and hypoxic pulmonary hypertension in mice by inhibition of Rho-kinase. *Am J Physiol Lung Cell Mol Physiol* 287:L656–L664
148. Guilluy C, Rolli-Derkinderen M, Tharaux PL, Melino G, Pacaud P, Loirand G (2007) Transglutaminase-dependent RhoA activation and depletion by serotonin in vascular smooth muscle cells. *J Biol Chem* 282:2918–2928
149. Guilluy C, Eddahibi S, Agard C, Guignabert C, Izikki M, Tu L, Savale L, Humbert M, Fadel E, Adnot S, Loirand G, Pacaud P (2009) RhoA and Rho kinase activation in human pulmonary hypertension: role of 5-HT signaling. *Am J Respir Crit Care Med* 179:1151–1158
150. Taraseviciene-Stewart L, Scerbavicius R, Choe KH, Cool C, Wood K, Tuder RM, Burns N, Kasper M, Voelkel NF (2006) Simvastatin causes endothelial cell apoptosis and attenuates

- severe pulmonary hypertension. *Am J Physiol Lung Cell Mol Physiol* 291:L668–L676
151. Toda N, Ayajiki K, Okamura T (2005) Nitric oxide and penile erectile function. *Pharmacol Ther* 106:233–266
  152. Bivalacqua TJ, Liu T, Musicki B, Champion HC, Burnett AL (2007) Endothelial nitric oxide synthase keeps erection regulatory function balance in the penis. *Eur Urol* 51:1732–1740
  153. Dean RC, Lue TF (2005) Physiology of penile erection and pathophysiology of erectile dysfunction. *Urol Clin North Am* 32:379–395
  154. Puetz S, Lubomirov LT, Pfitzer G (2009) Regulation of smooth muscle contraction by small GTPases. *Physiology (Bethesda)* 24:342–356
  155. Mills TM, Lewis RW, Wingard CJ, Linder AE, Jin L, Webb RC (2003) Vasoconstriction, RhoA/Rho-kinase and the erectile response. *Int J Impot Res* 15(Suppl 5):S20–S24
  156. Jin L, Burnett AL (2006) RhoA/Rho-kinase in erectile tissue: mechanisms of disease and therapeutic insights. *Clin Sci (Lond)* 110:153–165
  157. Chitaley K, Wingard CJ, Clinton Webb R, Branam H, Stopper VS, Lewis RW, Mills TM (2001) Antagonism of Rho-kinase stimulates rat penile erection via a nitric oxide-independent pathway. *Nat Med* 7:119–122
  158. Chitaley K, Webb RC, Dorrance AM, Mills TM (2001) Decreased penile erection in DOCA-salt and stroke prone-spontaneously hypertensive rats. *Int J Impot Res* 13(Suppl 5):S16–S20
  159. Burchardt M, Burchardt T, Baer L, Kiss AJ, Pawar RV, Shabsigh A, de la Taille A, Hayek OR, Shabsigh R (2000) Hypertension is associated with severe erectile dysfunction. *J Urol* 164:1188–1191
  160. Shabsigh R (2004) Testosterone therapy in erectile dysfunction. *Aging Male* 7:312–318
  161. Feldman HA, Johannes CB, Derby CA, Kleinman KP, Mohr BA, Araujo AB, McKinlay JB (2000) Erectile dysfunction and coronary risk factors: prospective results from the Massachusetts male aging study. *Prev Med* 30:328–338
  162. Kloner RA (2008) Erectile dysfunction: the new harbinger for major adverse cardiac events in the diabetic patient. *J Am Coll Cardiol* 51:2051–2052
  163. Buyukafsar K, Un I (2003) Effects of the Rho-kinase inhibitors, Y-27632 and fasudil, on the corpus cavernosum from diabetic mice. *Eur J Pharmacol* 472:235–238
  164. Wingard CJ, Johnson JA, Holmes A, Prikosh A (2003) Improved erectile function after Rho-kinase inhibition in a rat castrate model of erectile dysfunction. *Am J Physiol Regul Integr Comp Physiol* 284:R1572–R1579
  165. Vignozzi L, Morelli A, Filippi S, Ambrosini S, Mancina R, Luconi M, Mungai S, Vannelli GB, Zhang XH, Forti G, Maggi M (2007) Testosterone regulates RhoA/Rho-kinase signaling in two distinct animal models of chemical diabetes. *J Sex Med* 4:620–630 (discussion 631–632)
  166. Rajasekaran M, White S, Baquir A, Wilkes N (2005) Rho-kinase inhibition improves erectile function in aging male Brown-Norway rats. *J Androl* 26:182–188
  167. Jin L, Liu T, Lagoda GA, Champion HC, Bivalacqua TJ, Burnett AL (2006) Elevated RhoA/Rho-kinase activity in the aged rat penis: mechanism for age-associated erectile dysfunction. *FASEB J* 20:536–538
  168. Seasholtz TM, Wessel J, Rao F, Rana BK, Khandrika S, Kennedy BP, Lillie EO, Ziegler MG, Smith DW, Schork NJ, Brown JH, O'Connor DT (2006) Rho kinase polymorphism influences blood pressure and systemic vascular resistance in human twins: role of heredity. *Hypertension* 47:937–947